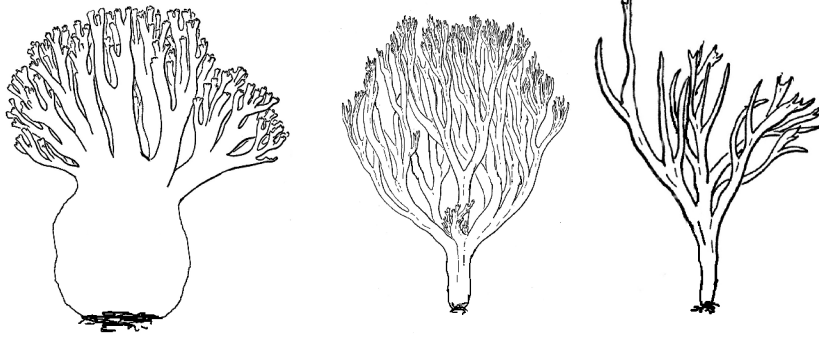


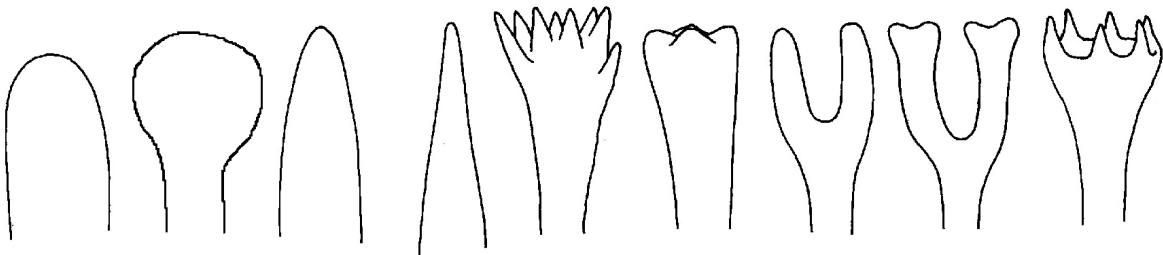
## Ramaria Description Sheet – ABRs Project

Use the sketches on this page where possible, but add extra remarks in the text entry areas on p2.



Or sketch it here:

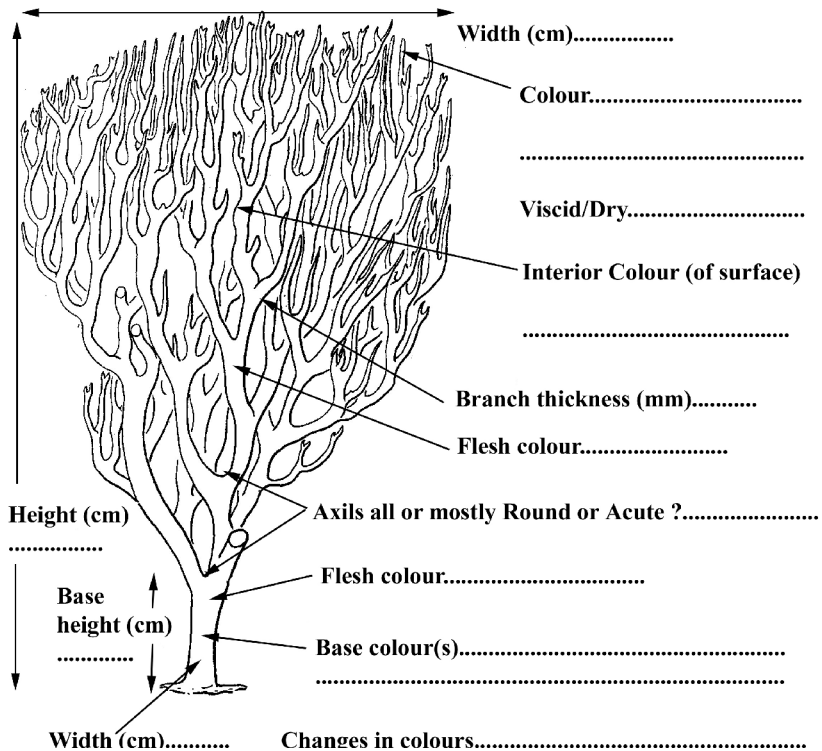
Tick the best fit. Use lines to alter the shape of the drawings above (or below) if needed.



Tick ***all*** the above apices that resemble your specimen. Record below if they are viscid/dry, and if possible, their colours when young, mature and old. Apices of a form second from the left are likely to be viscid when mature and often 'fused' together to form a surface. Apices may be intermediate between two samples and you can indicate this by joining with a line.

.....

.....



## Other Details

### Branches

Surfaces (smooth, wrinkled, grooved, etc.).....

Internodes (distance between forks..rapidly or slowly decreasing with height from stem).....

Orientation(vertically, branching 'radially', horizontal then vertical, etc.).....

### Stem

Surface (smooth, velvety, fluffy, hairy, etc.).....

Shape (bulbous, slender, etc.).....

Aborted branches present or absent.....

Single, fasciculate(several stems clustered closely), single stem composed of several fused stems, etc..

**Colour changes** (when bruised or cut..these may be slow and may occur anywhere on the fungus).....

### Growth habit

Solitary, clumped, gregarious, occasional, etc...give further details.....

### Substrate

Humus, rotting wood, treefern base, soil, etc.....

**Mycelial mat and Rhizomorphs** (Forest species may produce a white carpet of mycelium and rhizomorphs – white strands like thin cotton) present/absent for each.....

**Habitat:** forest, woodland, heath, etc...give further details.....

**Photographs** (circle): slide photograph digital (We will contact you for digital transmission)

**Odour/Taste(if noted):** .....

**Spore print colour (see note below):**.....

**Date:**...../...../..... **Location:**..... **State:**.....

**GPS (Type if known, give lat/long if possible):**.....

**Collector(s):**.....

## Suggestions on Using Description Sheets and Preparing Material

- 1. General Items** Fruiting bodies may not conform to the sample drawings perfectly; modify the drawings, add your own drawings and give explanatory notes. Anything is useful. The best colour reference chart for easy use is that produced by the Royal Botanic Gardens Edinburgh...it costs about \$1.00 per copy (plus postage). It has been specifically developed for use with fungi and is very simple to apply.
- 2. Photographs**

Colour photographs are vital for good identification with this genus. Photo slides or digital images are best, but colour photographs can be used. Take at least two (preferably three) photos per specimen: an “in situ” shot (with the fungus undisturbed) from a side distance of 30–50 cm at an angle permitting good feature display; and a second side shot (with distance of 30–40 cm) after removal from the substrate. A 5 cm scale or a 20 cent coin can be included, but keep coin or scale clearly separate from the fungus. Please try to take a third shot to obtain excellent, close-up resolution of the apices – their shapes and colours are critically important. If there is a quantity of the fungus, a fourth longer distance shot showing the fungus in its habitat situation would be very much appreciated. For digital photographs, please include your name, address and email address. We will make arrangements to obtain copies of the jpegs/images from you.
- 3. Colour Changes**

Some species change colour on bruising or the flesh may alter colour after cutting; this may be immediate or over perhaps half an hour. Please record the time taken and the colour changes.
- 4. Axils**

The branch axils are sometimes very difficult to describe. They are usually rounded or V-shaped (acute). Sometimes the branches are very acute, but the ultimate junction still remains rounded. If there is a mixture of both, indicate this on the sheet.
- 5. Spore Prints**

Spore prints are often produced in containers during transport from the field. A good way to take advantage of this is to put some white paper under the fungus and the print is often ready by the time you have the fungus back home or at the lab. Otherwise, place the fungus on white paper, cover it with a plastic or glass bowl and leave overnight. If the fungus is sporulating, a good spore print should be present in the morning. This can be allowed to dry, folded and stored in an envelope with the specimen. If you have a colour chart, indicate the colour chip that describes the print colour.
- 6. Rhizomorphs – a very special requirement !!!!!!!!!!!!!**

If present (not all species have them), rhizomorphs appear as white, “cotton-thread-like” strands attached to the base of the fruiting body. It is absolutely vital that they be collected with the specimen (preferably still attached) because successful ID's depend on their microscopic structures.
- 7. Drying the Collections**

Dry material in the air..the best method is to use a fruit dryer (Vacola set to about 40 degrees C) but any warm air current can be used. Drying should be done as fast as convenient to prevent deterioration of the specimens.
- 8. Locations**

If you have a GPS unit, give the lat/long and the name of the location...If no GPS unit is held, give the name of the site plus the name of the nearest reasonably sized town or village. If you can, provide essential details (distances, features, signposts etc.) so the site can be re-located exactly.
- 9. Habitat Details** For the habitat, a general idea such as rainforest, wet eucalypt forest, dry eucalypt-casuarina woodland, beech forest, heath, etc. will be very helpful. If you know precisely the type of trees under which the specimen is growing, include this as well...we know very little about the ecology of these fungi.

## 10.Substrates

Many ground substrates will be simply “on soil amongst litter, etc.”. If possible, indicate the type of soil, eg. Basaltic, granitic, sandy, black loam, etc. This may be very difficult and impossible under some circumstances...It's nice to have if we can get it.

## 11.Adding your own Notes

If you want to add additional notes on your own topics...feel absolutely free, we'd really like them.

## 12.Collection Details

Don't forget to put down who collected the fungus and when. We want to give credit to the collectors.

## 13.Posting Instructions for Collections

When sending dried material through the post, ensure that the material is sent in crush proof boxes. Dried coralloid fungi are *extremely* fragile and putting them into padded bags only, almost ensures that they will arrive here crushed. If you must put them into small packets, then post the packets inside a crush proof box with ample padding so that they will not bounce around in transport.

## 14.Contacting Us

Please don't hesitate to contact us by phone or email if you need to ask any other questions.

### Special Note....Chemical Tests

For those with extra laboratory equipment, the coralloid fungi have been tested with a number of simple reagents and the fresh tissues often exhibit colour changes under these reagents. The most common and easily obtainable reagents used are:

10% solution of ferrous sulphate  
Melzers reagent (or Lugol's iodine would probably suffice)  
2% Hydrochloric acid  
2% potassium hydroxide

If you can get test solutions of these, the method is simply to put a drop of the solution on a freshly cut surface and note/record the resulting colour change, if any...You should also try the solution on the stem or branch surface as well.

*And finally, a very big thankyou for generously agreeing to help us so much.*

Tony Young & Nigel Fechner.

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